SCIENTIFIC **Reports**

Received: 20 December 2016 Accepted: 5 June 2017 Published online: 20 July 2017

OPEN Comparative transcriptome analysis of the floral transition in *Rosα chinensis* 'Old Blush' and *R*. odorata var. gigantea

Xuelian Guo, Chao Yu, Le Luo, Huihua Wan, Yushu Li, Jia Wang, Tangren Cheng, Huitang Pan & Qixiang Zhang

The floral transition is a crucial developmental event, but little is known about the underlying regulatory networks in seasonally and continuously flowering roses. In this study, we compared the genetic basis of flowering in two rose species, Rosa chinensis 'Old Blush', which flowers continuously, and R. odorata var. gigantea, which blooms in early spring. Gene ontology (GO) terms related to methylation, light reaction, and starch metabolism were enriched in R. odorata var. gigantea and terms associated with sugar metabolism were enriched in R. chinensis 'Old Blush' during the floral transition. A MapMan analysis revealed that genes involved in hormone signaling mediate the floral transition in both taxa. Furthermore, differentially expressed genes (DEGs) involved in vernalization, photoperiod, gibberellin (GA), and starch metabolism pathways converged on integrators, e.g., LFY, AGL24, SOC1, CAL, and COLs, to regulate the floral transition in R. odorata var. gigantea, while DEGs related to photoperiod, sugar metabolism, and GA pathways, including COL16, LFY, AGL11, 6PGDH, GASA4, and BAM, modulated the floral transition in R. chinensis 'Old Blush.' Our analysis of the genes underlying the floral transition in roses with different patterns of flowering provides a basis for further functional studies.

The floral transition is an important developmental event orchestrated by external and developmental stimuli; for example, photoperiod, vernalization, autonomous, aging, gibberellin (GA), and sugar metabolism pathways¹. ² interact to activate or inhibit the floral transition via a small set of floral integrators, including FLOWERING LOCUS T (FT), CONSTANS-LIKE (COL), SUPPRESSOR OF OVEREXPRESSION OF CO1 (SOC1), FLOWERING LOCUS C (FLC), and floral meristem identity genes, e.g., LEAFY (LFY), APETALA1 (AP1), AGAMOUS-LIKE (AGL), CAULIFLOWER (CAL), and SEPALLATA (SEP).

Considerable progress has been made toward identifying the molecular networks underlying the floral transition in the annual plant Arabidopsis thaliana³. However, relatively little is known about the regulation of the floral transition in perennial plants. In perennials, only a portion of buds have competence to complete the phase transition from vegetative to reproductive growth in inductive conditions, while others remain in vegetative growth, indicating the ability to revert back from reproductive to vegetative growth. In temperate regions, most perennials require a period of winter cold (vernalization) to induce the floral transition and show seasonal flowering. However, a small number of perennials, such as Rosa chinensis and Fragaria vesca, can flower continuously in advantageous environmental conditions. The duration of flowering is crucial for the value of ornamental plants, such as modern roses and chrysanthemums.

Some progress has been made in understanding flowering perennials. Vernalization and sugar metabolism, which interacts with ABA signaling and aging pathways, are key factors inducing the floral transition in Malus × domestica^{4,5}. The photoperiod-mediated CO/FT modules function as activators in the induction of the floral transition in chrysanthemums and poplars^{6,7}. Different from other species in the genus Camellia, the floral transition in C. azalea occurs in the spring; it is regulated by low temperatures, photoperiod, and hormone

Beijing Key Laboratory of Ornamental Plants Germplasm Innovation & Molecular Breeding, National Engineering Research Center for Floriculture, Beijing Laboratory of Urban and Rural Ecological Environment, Key Laboratory of Genetics and Breeding in Forest Trees and Ornamental Plants of Ministry of Education, School of Landscape Architecture, Beijing Forestry University, Beijing, 100083, China. Xuelian Guo and Chao Yu contributed equally to this work. Correspondence and requests for materials should be addressed to Q.Z. (email: zqxbjfu@126.com)